

RNA Secondary Structures

1st EVBC WinterSchool

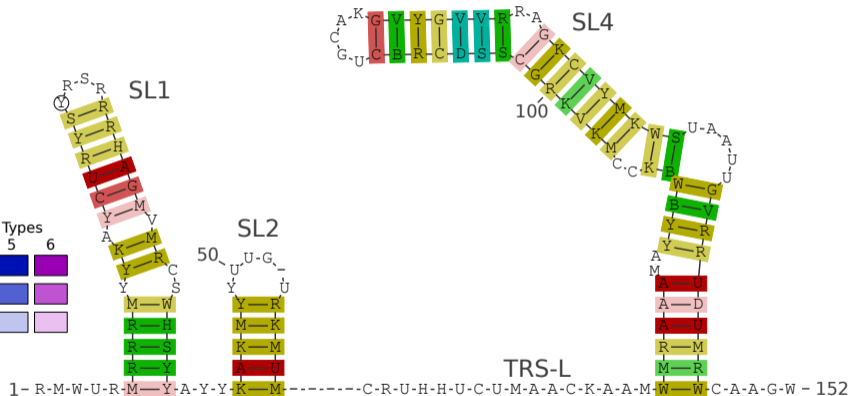
Jörg Fallmann, Kevin Lamkiewicz

9 – 13.03.2018

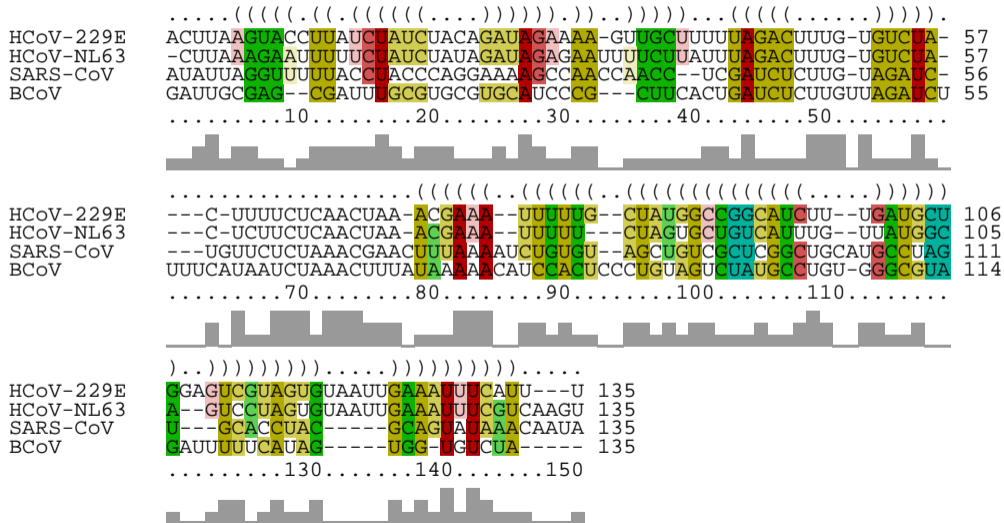


Alignments and compensatory mutations

UNCONSERVED SEQUENCE, CONSERVED STRUCTURE



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COMPENSATORY MUTATIONS IN SECONDARY STRUCTURES

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Compensatory mutations underline the importance of a specific secondary structure.

But be careful!

If we're assuming a uniform mutation rate, every third pair of mutations is a compensatory mutation.

A	U
A	A
A	C
A	G
C	A
C	C
C	G
C	U

A	U
G	A
G	C
G	G
G	U
U	A
U	C
U	G
U	U

RNA-RNA Long-Range Interactions

WHY LRIs?

- ▶ Interaction spans distances between a few hundred and several thousands of nucleotides
- ▶ few are described in positive stranded RNA viruses
- ▶ often located in loop regions (bulges, hairpins, ...)

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- ▶ often located in loop regions (bulges, hairpins, ...)
⇒ pseudo-knots!
- ▶ LRIs may play a very important role in viral replication

HOW TO CALCULATE LRIS

Approach I

- ▶ RNA duplex
- ▶ RNAplex
- ▶ RNAhybrid

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Approach III

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- ▶ IntaRNA

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Approach IV

- ▶ inteRNA
- ▶ inRNAs

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Approach V

- ▶ PETcofold
- ▶ RNAaliduplex

LRISCAN

Prediction of conserved long-range RNA-RNA interactions in full viral genomes, 2016. M. Fricke, M. Marz

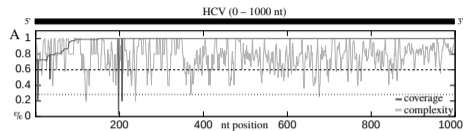
LRISCAN

Prediction of conserved long-range RNA-RNA interactions in full viral genomes, 2016. M. Fricke, M. Marz

⇒ LRIScan

How does LRIs work and how do I use it?

WORKFLOW OF LRISCAN



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Coverage of an alignment

Relative number of sequences that do not have a gap on a specific position.

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$$C_i = \frac{1}{m} \sum_{k=1}^m \frac{|\delta(a_{i \dots i+s-1}^k)|}{|(a_{i \dots i+s-1}^k)|}$$

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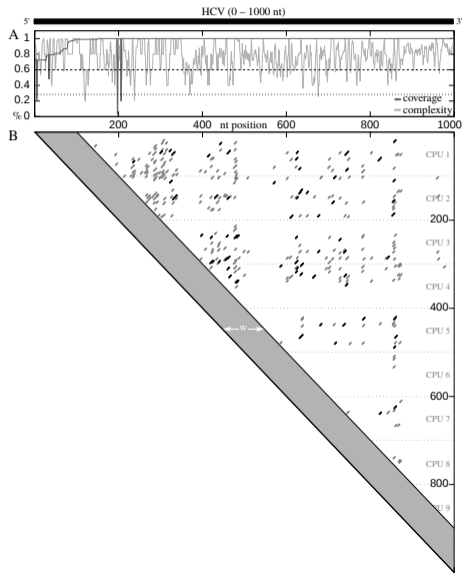
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Complexity of the alignment

$$C_i = \frac{1}{m} \sum_{k=1}^m \frac{|\delta(a_{i \dots i+s-1}^k)|}{|(a_{i \dots i+s-1}^k)|}$$

$$\delta(\text{CCUUUGGAAA}) = \text{CUGA}$$

WORKFLOW OF LRISCAN – STEP 2

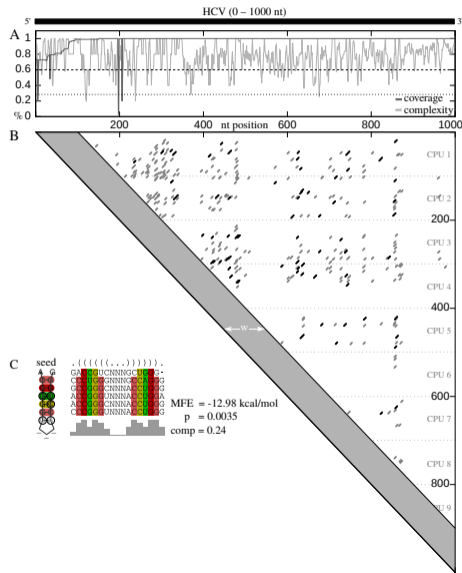


FINDING SEEDS

$$S_{i,j} = (S_{i-1,j+1} + 1) \cdot \Pi_{ij} \cdot \Phi_{ij}$$

- ▶ Π_{ij} : do at least t percent of the input sequence form the basepair (i, j) ?
- ▶ Φ_{ij} : do both alignment columns A_i and A_j meet the coverage threshold?

WORKFLOW OF LRISCAN – STEP 3



SEED SCORING

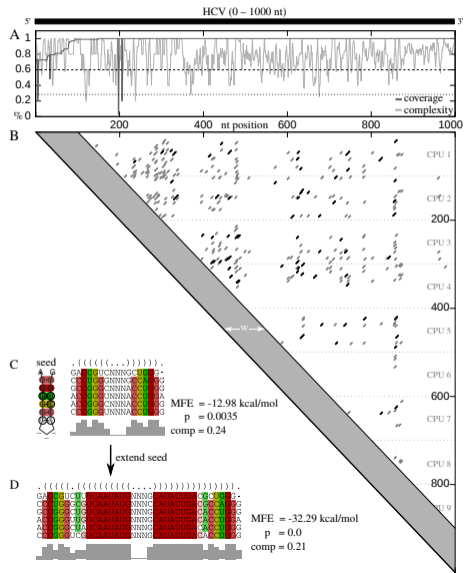
- ▶ z-Score analysis for each seed to measure reliability
- ▶ compensatory score τ

$$\tau = \frac{\sum_b (u \cdot h)}{6 \cdot |b| \cdot k}$$

with:

- ▶ u : number of different base-pair types
- ▶ h : number of incompatible base-pairs

WORKFLOW OF LRISCAN – STEP 4



SEED EXTENSION

- ▶ each seed is extended 10 nts at the 5' (and 3' respectively)
- ▶ calculate MFE with RNAalifold
 - ▶ hard constraints for seed region
 - ▶ soft constraints for extension, such that intermolecular interactions are formed

LRIsScan Results and Output

LRISCAN USAGE

```
1 $> ./LRIsScan.rb -c 2 -f <ALIGNMENT> -o <OUTPUT>
```

- ▶ tabular output in .tsv format
- ▶ table and figures in .html
- ▶ all figures are also stored in the `ps/` directory

Example Data Set

QUESTIONS

We want to analyze the human coronavirus 229e (HCoV229e).

- ▶ Are there conserved structures in the 5' UTR?
- ▶ Are there conserved structures in the 3' UTR?
- ▶ Can we detect long-range interactions within the Coronaviruses?
- ▶ The virus with the sequence of `hcov229e_mutation.fasta` does not replicate as well as the wildtype. Our wet-lab collaboration partner assumes there might be something odd in the 5' UTR.
- ▶ Extract the 5'UTR of the `hcov229e_5utr_mutation.fasta`. How does the secondary structure behave?
- ▶ There might be a pseudoknot in the 3'UTR. Do you have any ideas how to confirm this hypothesis?
- ▶ ...

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Note:

If you have your own ideas on what to analyze, feel free to do so. These are just some example questions. Furthermore, if you want to include more related viruses to your analyses, you're welcome to do so as well.